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(54) Title: METHOD OF DETECTING PRIONS IN A SA	AMPLI	E AND TRANSGENIC ANIMAL USED FOR SAME
(57) Abstract		
The invention includes an artificial Pr2 gene, a transgused to detect the presence of pathogenic prions in a sample		nimal containing the gene, and an assay wherein the transgen's inimal is agnose a cause of death.

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METHOD OF DETECTING PRIONS IN A SAMPLE AND TRANSGENIC ANIMAL USED FOR SAME

Government Rights

The United States Government may have certain rights in this application pursuant to Grant Nos. NS14069, 5 AG02132, NS22786, AG08967 and AG10770 awarded by the National Institutes of Health.

Field of the Invention

This invention relates generally to chimeric genes, 10 methods of as lying and to transgenic animals used in such assays. More specifically, this invention relates to artificial and chimeric PrP genes, assaying samples for pathogenic prions, and to transgenic mice containing an artificial or chimeric PrP gene.

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Background of the Invention

Prions are infectious pathogens that cause central nervous system spongiform encephalopathies in humans and animals. Prions are distinct from bacteria, viruses and The predominant hypothesis at present is that no nucleic acid component is necessary for infectivity of Further, a prion which infects one prion protein. species of animal (e.g., a human) will not infect another (e.g., a mouse).

A major step in the study of prions and the diseases that they cause was the discovery and purification of a protein designated prion protein ("PrP") [Bolton et al., Science 218:1309-11 (1982); Prusiner et al., Biochemistry 21:6942-50 (1982); McKinley et al., Cell 35:57-62 Complete prion protein-encoding genes have (1983)]. since been cloned, sequenced and expressed in transgenic Prpc is encoded by a single-copy host gene [Basler et al., Cell 46:417-28 (1986)] and is normally found at the outer surface of neurons. 35 hypothesis is that prion diseases result from conversion of PrPc into a modified form called PrPsc. However, the

actual biological or physiological function of PrP is not known.

It appears that the scrapie isoform of the prion protein (PrPsc) is necessary for both the transmission and 5 pathogenesis of the transmissible neurodegenerative diseases of animals and humans. See Prusiner, S.B., "Molecular biology of prion disease," Science 252:1515-1522 (1991). The most common prion diseases of animals are scrapie of sheep and goats and bovine spongiform 10 encephalopathy (BSE) of cattle (Wilesmith, J. and Wells, Microbiol. Immunol. 172:21-38 (1991)]. diseases of humans have been identified: (1) kuru, (2) Creutzfeldt-Jakob Disease (CJD), (3) Gerstmann-Strassler-Scheinker Disease (GSS), and (4) fatal familial 15 insomnia (FFI) [Gajdusek, D.C., Science 197:943-960 (1977); Medori, et al., N. Engl. J. Med. 326:444-449 (1992)]. The presentation of human prion diseases as sporadic, genetic and infectious illnesses initially posed a conundrum which has been explained by the 20 cellular genetic origin of PrP.

Most CJD cases are sporadic, but about 19-15% are inherited as autosomal dominant disorders that are caused by mutations in the human PrP gene [Hsiao et al., Neurology 40:1820-1827 (1990); Goldfarb et al., Science 25 258:806-808 (1992); Kitamoto et al., Proc. R. Soc. Lond. (In press) (1994)]. Iatrogenic CJD has been caused by human growth hormone derived from cadaveric pituitaries as well as dura mater grafts [Brown et al. Lancet 340:24-27 (1992)]. Despite numerous attempts to link CJD to an 30 infectious source such as the consumption of scrapie infected sheep meat, none has been identified to date [Harries-Jones et al., J. Neurol. Neurosurg. Psychiatry 51:1113-1119 (1988)] except in cases of iatrogenically induced disease. On the other hand, kuru, which for many 35 decades devastated the Fore and neighboring tribes of the New Guinea highlands, is believed to have been spread by infection during ritualistic cannibalism [Alpers, M.P.,

Slow Transmissible Diseases of the Nervous System, Vol. 1, S.B. Prusiner and W.J. Hadlow, eds. (New York: Academic Press), pp. 66-90 (1979)].

The initial transmission of CJD to experimental 5 primates has a rich history beginning with William Hadlow's recognition of the similarity between kuru and scraple. In 1959, Hadlow suggested that extracts prepared from patients dying of kuru be inoculated into non-human primates and that the animals be observed for 10 disease that was predicted to occur after a prolonged incubation period [Hadlow, W.J., Lancet 2:289-290 (1959)]. Seven years later, Gajdusek, Gibbs and Alpers demonstrated the transmissibility of kuru to chimpanzees after incubation periods ranging form 18 to 21 months 15 [Gajdusek et al., Nature 209:794-796 (1966)]. similarity of the neuropathology of kuru with that of CJP [Klatzo et al., Lab Invest. 8:799-847 (1959)] prompted similar experiments with chimpanzees and transmissions of disease were reported in 1968 [Gibbs, Jr. et al., Science 20 161:388-389 (1968)]. Over the last 25 years, about 300 cases of CJD, kuru and GSS have been transmitted to a variety of apes and monkeys.

The expense, scarcity and often perceived inhumanity of such experiments have restricted this work and thus limited the accumulation of knowledge. While the most reliable transmission data has been said to emanate from studies using non-human primates, many cases of human prion disease have been transmitted to rodents but apparently with less regularity [Gibbs, Jr. et al., Slow Transmissible Diseases of the Nervous System, Vol. 2, S.B. Prusiner and W.J. Hadlow, eds. (New York: Academic Press), pp. 87-110 (1979); Tateishi et al., Prion Diseases of Humans and Animals, Prusiner et al. eds. (London: Ellis Horwood), pp. 129-134 (1992)].

The infrequent transmission of human prior disease to rodents has been cited as an example of the "species barrier" first described by Pattison in his studies of

passaging the scrapic agent between sheep and rodents [Pattison, I.H., NINDB Monograph 2, D.C. Gajdusek, C.J. Gibbs Jr. and M.P. Alpers, edr. (Washington, D.C.: U.S. Government Printing), pp. 249-257 (1965)]. In those investigations, the initial passage of prions from one species to another was associated with a prolonged incubation time with only a few animals developing illness. Subsequent passage in the same species was characterized by all the animals becoming ill after greatly shortening incubation times.

The molecular basis for the species barrier between Syrian hamster (SHa) and mouse was shown to reside in the sequence of the PrP gene using transgenic (Tg) mice [Scott et al., Cell 59:847-857 (1989)]. SHaPrP differs 15 from MoPrP at 16 positions out of 254 amino acid residues [Basler et al., Cell 16:417-428 (1986); Locht et al., Proc. Natl. Acad. Sci. USA 83:6372-6376 (1986)]. mice expressing SHaPrP had abbreviated Tq(SHaPrP) incubation times when inoculated with SHa prions. 20 similar studies were performed with mice expressing the human, or ovine PrP transgenes, the species barrier was not abrogated, i.e., the percentage of animals which became infected were unacceptably low and the incubation times were unacceptably long. Thus, it has not been 25 possible to use transgenic animals (such as mice containing a PrP gene of another species) to reliably test a sample to determine if that sample is infected The seriousness of the health risk with prions. resulting from the lack of such a test is exemplified 30 below.

More than 45 young adults previously treated with HGH derived from human pituitaries have developed CJD [Koch et al., N. Engl. J. Med. 313:731-733 (1985); Brown et al. Lancet 340:24-27 (1992); Fradkin et al., JAMA 265:880-884 (1991); Buchanan et al., Br. Med. J. 302:824-828 (1991)]. Fortunately, recombinant HGH is now used, although the seemingly remote possibility has been raised

that increased expression of wtPrP stimulated by high HGH might induce prion disease [Lasmezas et al., Biochem. Biophys. Res. Commun. 196:1163-1169 (1993)]. HGH prepared from pituitaries was contaminated with 5 prions is supported by the transmission of prion disease to a monkey 66 months after inoculation with a suspect lot of HGH [Gibbs, Jr. et al., N. Engl. J. Med. 328:358-359 (1993)]. The long incubation times associated with prion diseases will not reveal the full extent of iatrogenic CJD for decades in thousands of people treated with HGH worldwide. Iatrogenic CJD also appears to have developed in four infertile women treated contaminated human pituitary-derived gonadotrophin hormone [Healy et al., Br. J. Med. 307:517-518 (1993); 15 Cochius et al., Aust. N.Z. J. Med. 20:592-593 (1990); Cochius et al., J. Neurol. Neurosurg. Psychiatry 55:1094-1095 (1992)] as well as at least 11 patients receiving dura mater grafts [Nisbet et al., J. Am. Med. Assoc. 261:1118 (1989); Thadani et al., J. Neurosurg, 69:766-769 20 (1988); Willison et al., J. Neurosurg. Psychiatric 54:940 (1991); Brown et al. Lancet 340:24-27 (1992)]. cases of iatrogenic CJD underscore the need for screening pharmaceuticals that might possibly be contaminated with prions.

25 Recently, two doctors in France were charged with involuntary manslaughter of a child who had been treated with growth hormones extracted from corpses. The child developed Creutzfeldt-Jakob Disease. (See New Scientist, July 31, 1993, page 4). According to the Pasteur 30 Institute, since 1989 there have been 24 reported cases of CJD in young people who were treated with human growth hormone between 1983 and mid-1985. Fifteen of these children have died. It now appears as though hundreds of children in France have been treated with growth hormone 35 extracted from dead bodies at the risk of developing CJD (see New Scientist, November 20, 1993, page 10.) In view of such, there clearly is a need for a convenient, cost-

effective assay for desting sample materials for the presence of prions which cause CJD. The present invention offers such an assay.

Summary of the Invention

The invention includes an artificial PrP gene, a transgenic animal containing the gene, and assay methodology which uses the transgenic animal to detect pathogenic prions in a sample. The artificial gene 10 includes a sequence such that when it is inserted into the genome of an animal (such as a mouse), the animal is rendered susceptible to infection with prions which normally would infect only a specific species of genetically diverse animal (such as human, cow or sheep). 15 The artificial PrP gene may be comprised of a completely artificial polynucleotide sequence. Alternatively, the artificial gene may be comprised of the codon sequence of a first animal with one or more codon substitutions being the substitutions preferably made wherein are 20 corresponding PrP gene codons from a genetically diverse animal with the proviso that not all of the different codons are replaced by codons of the genetically diverse animal.

Pathogenic prions in a sample can be detected by injecting the sample to be tested into a transgenic In one preferred example the mouse genome includes a chimeric PrP gene which gene includes a portion of a gene of the animal (e.g. human) in danger of infection from prions in the sample. For example, is fatal Disease (CJD) 30 Creutzfeldt Jakob neurodegenerative disease of humans caused by prions. Preferred transgenic (Tg) mice disclosed herein express a chimeric prion protein (PrP) in which a segment of mouse (Mo) PrP was replaced with the corresponding human 35 (Hu) PrP sequence. The chimeric PrP designated MHu2MPrP, differs from MoPrP by 9 amino acids between residues 96 and 167. All of the Tg(M:lu2MPrP) mice injected with

human prions developed neurologic disease. More specifically, the transgenic mice of the invention developed the disease -200 days after inoculation with brain homogenates from three CJD patients. 5 inoculated with CJD prions, MHu2MPrPsc was formed; in contrast MoPrPsc was produced if Mo prions were The patterns of MHu2MPrPsc and Moprpsc inoculated. accumulation in the brains of Tg(MHu2M) mice were different. About 10% of Tg(HuPrP) mice expressing HuPrP 10 and Non-Tg mice developed neurologic disease >500 days after inoculation with CJD prions. The different susceptibilities of Tg(HuPrP) and Tg(MHu2M) mice to human prions indicates that additional species specific factors are involved in prion replication. Tg(MHu2MPrP) mice 15 disclosed herein are useful in the diagnosis, prevention and treatment of human prion diseases. Transgenic mice containing the chimeric prion protein gene which includes a portion, but not all, of the PrP gene of an animal from a different genus can be used to test samples for prions 20 which might infect such animals. The transgenic mice disclosed herein consistently develop the adverse effects of such prions in a relatively short time and are substantially cheaper and easier to maintain than are currently used primate models.

An object of the invention is to provide a gene which may be artificial or chimeric which gene when inserted into the genome of one animal (e.g., a mouse) will render the mammal susceptible to infections from prions which naturally only infect a genetically diverse 30 mammal, e.g., human, bovine or ovine.

Another object of the invention is to provide an assay for the detection of prions in a sample.

Another object is to provide an artificial PrP gene wherein one or more codons (preferably 1-50 codons) of 35 the PrP gene of a first animal (e.g. a mouse) is replaced with codons of the PrP gene of a genetically diverse animal (e.g. a human, cow or sheep) in a manner so as to

render the first animal susceptible to infection with prions which normally infect only the genetically diverse animal.

Another object is to provide a chimeric gene 5 comprised of codons encoding the C- and N- terminus of one species of mammal and middle codons of another species of mammal.

Another object of the invention is to provide a transgenic mammal such as a mouse which includes a chimeric PrP gene which gene includes a portion of the PrP gene from another animal, such as a human, cow, or sheep.

An advantage of the present invention is that the transgenic mouse can be used to assay for the presence of prions in a sample in a manner which is substantially faster, more efficient and cheaper than presently available assay methods.

Another advantage is that transgenic mice inoculated with prions of numans can be used as test animals for testing drugs for efficacy in the treatment of humans suffering from diseases resulting from infection with prions.

Another advantage is that the transgenic mice can detect prions in a sample at very low levels, e.g., 25 1 part per million, and even as low as 1 part per billion.

Still another advantage is that the mice provide an assay which is highly accurate, i.e., does not provide false positives and consistently determines the presence of prions.

Yet another advantage is that by increasing the copy number of the gene of the invention in a transgenic mammal, the incubation time for prion caused disease is decreased.

A feature of the present invention is that the transgenic mice injected with a sample containing pathogenic prions will consistently develop the disease

effects of the prions within a relatively short time, e.g. about 200 days \pm 50 days after injection or less.

Another feature is that an artificial gene of the invention preferably contains codons of the PrP gene of a first animal (such as a mouse) with some (but not all) of the codons which differ from the mouse and a second genetically diverse mammal (such as a human) replacing codons of the first mammal at the same relative positions.

These and other objects, advantages, and features of the invention will become apparent to those persons skilled in the art upon reading the details of the chimeric gene, assay method, and transgenic mouse as more fully described below.

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Brief Description of the Drawings

Figure 1 is a schematic drawing showing the construction of a chimeric MHu2M gene and a transgenic mouse containing same;

20 Figure 2 is a schematic view of a portion of PrP proteins showing the differences between a normal, wild-type numan PrP protein and a normal, wild-type mouse PrP protein;

Figure 3 shows the amino acid sequence of mice PrP 25 along with specific differences between mice PrP and human PrP;

Figure 4 shows the amino acid sequence of mouse PrP and specifically shows differences between mouse PrP and bovine PrP; and

30 Figure 5 shows the amino acid sequence of mouse PrP and specifically shows differences between mouse PrP and ovine PrP.

Detailed Description of Preferred Embodiments

Before the present chimeric gene, assay methodology and transgenic mice used in the assay are described, it is to be understood that this invention is not limited to

particular assay methods, chimeric and artificial genes or transgenic mice described, as such methods, chimeric genes and mice may, of course, vary. It is also to be understood that the terminology used herein is for the 5 purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims.

defined otherwise, all Unless technical 10 scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the 15 present invention, the preferred methods and materials are now described. All publications mentioned herein are incorporated herein by reference to disclose and describe the methods and/or materials in connection with which the publications are cited.

The term "prion" shall mean an infectious particle known to cause diseases (spongiform encephalopathies) in humans and animals. The term "prion" is a contraction of the words "protein" and "infection" and the particles are comprised largely if not exclusively of PrPsc molecules 25 encoded by a Prp gene. Prions are distinct from bacteria, viruses and viroids. Known prions include those which infect animals to cause scrapie, transmissible, degenerative disease of the nervous system sheep and goats as well as bovine spongiform 30 encephalopathies (BSE) or mad cow disease and feline spongiform encephalopathies of cats. Four prion diseases known to affect humans are (1) kuru, (2) Creutzfeldt-Jakob Disease (CJD), (3) Gerstmann-Strassler-Scheinker Disease (GSS), and (4) fatal familial insomnia (FFI). As 35 used herein prion includes all forms of prions causing all or any of these diseases or others in any animals

used - and in particular in humans and in domesticated farm animals.

The term "PrP gene" refers generally to any gene of any species which encodes any form of a prion protein.

The term "artificial PrP gene" is used herein to 5 encompass the term "chimeric PrP gene" as well as other recombinantly constructed genes which whe included in the genome of one animal (e.g., a mouse) will render the susceptible to infection from prions which mammal naturally only infect a genetically diverse mammal, e.g., human, bovine or ovine. In general, an artificial gene will include the codon sequence of the PrP gene of the mammal being genetically altered with one or more (but not all, and generally less than 50) codons of the natural sequence being replaced with a different codon preferably a corresponding codon of a genetically diverse mammal (such as a human). The genetically altered mammal being used to assay samples for prions which only infect the genetically diverse mammal. Examples of artificial genes are mouse PrP genes encoding the sequence as shown 20 in Figures 3, 4 and 5 with one or more different replacement codons selected from the codons shown in these Figures for humans, cows and sheep replacing mouse codons at the same position, with the proviso that not 25 all the mouse codons are replaced with differing human, cow or sheep codons.

The terms "chimeric gene," "chimeric PrP gene" and the like are used interchangeably herein to mean un artificially constructed gene containing the codons of a 30 first animal such as a mouse with one or more of the codons being replaced with corresponding codons from a genetically diverse animal such as a human, cow or sheep. In one specific example the chimeric gene is comprised of the starting and terminating sequence (i.e., N- and Cterminal codons) of a PrP gene of a mammal of a first species (e.g. a mouse) and also containing a nucleotide sequence of a corresponding portion of a PrP gene of a

mammal of a second species (e.g. a human). A chimeric gene will, when inserted into the genome of a mammal of the first species, render the mammal susceptible to infection with prions which normally infect only mammals 5 of the second species. The preferred chimeric gene disclosed herein is MHu2M which contains the starting and terminating sequence of a mouse PrP gene and a nonterminal sequence region which is replaced with a corresponding human sequence which differs from a mouse 10 PrP gene in a manner such that the protein expressed thereby differs at nine residues.

terms "Lusceptible to infection" "susceptible to infection by prions" and the like are used interchangeably herein to describe a transgenic 15 mammal of the invention which has an 80% or greater, preferably 98% or greater, and most preferably a 100% chance of developing a disease if inoculated with prions. The terms are used to describe a transgenic animal of the invention such as a transgenic mouse Tg(MHu2M) which, 20 without the chimeric PrP gene, would not be susceptible to infection with a human prion (less than 20% chance of infection) but with the chimeric gene is susceptible to infection with human prions (80% to 100% chance of infection).

The term "incubation time" shall mean the time from inoculation of an animal with a prion until the time when the animal first develops detectable symptoms of disease resulting from the infection. A reduced incubation time is one year or less, preferable about 200 days ± 50 days 30 or less, more preferably about 50 days ± 20 days days or less.

CJD for herein include Abbreviations used Creutzfeldt-Jakob Diseaue; GSS for Gerstmann-Strassler-Scheinker Disease; FFI for fatal familial insomnia; PrPsc 35 for the scrapie isoform of the prion protein; MoPrP for a mouse prion protein; SHa for a Syrian hamster; BSE for bovine spongiform encephalopathy; CNS for central nervous system; MHu2M for a chimeric mouse/human PrP gene wherein a region of the mouse PrP gene is replaced by a corresponding human sequence which differs from mouse PrP at 9 codons; Tg(MHu2M) mice are transgenic mice of the invention which include the MHu2M gene;

CNS for central nervous system;

CJD for Creu-zfeldt-Jakob Disease;

GSS for Gerstmann-Strassler-Scheinker Disease;

FFI for fatal familial insomnia;

10 Hu for human;

HuPrP for a human prion protein;

Mo for mouse;

SHaPrP for a Syrian hamster prion protein;

Tq for transquaic;

15 Tg(SHaPrP) for a transgenic mouse containing the PrP gene of a Syrian hamster,

Tg(HuPrP) for transgenic mice containing the complete human PrP gene;

Tg(ShePrP) for transgenic mice containing the complete sheep PrP gene;

Tg(BovPrP) for transgenic mice containing the complete cow PrP gene;

MoPrP^{sc} for the scrapie isoform of the mouse prion protein;

25 MHu2MPrPsc for a chimeric gene wherein a portion of the scrapie isoform of the human PrP gene is fused into the scrapie isoform of the mouse PrP gene.

Prpcn for the CJD isoform of a PrP gene;

Prn-p^{0/0} for ablation of both alleles of the MoPrP gene;

30 Tg(SHaPrP*'0)81/Prn-p0'0 for a particular line (81) of transgeneic mice expressing SHaPrP, +/0 indicates heterozygous;

General Aspects of the Invention

35 The present invention includes several aspects including: (1) an artific in gene comprised of codon sequences which when inserted into the genome of a first

animal (e.g. a mouse or rat) will render the animal susceptible to infection with prions which normally infect only a genetically diverse animal (e.g. a human, cow or sheep), thereby including genes wherein 1 to 50 5 codons of a naturally occurring PrP gene of a first animal are replaced with corresponding codons of a genetically diverse animal; (2) a chimeric gene which gene is comprised of the PrP sequence of a gene of a mammal of a first species which gene has been modified to 10 include a corresponding segment of a PrP gene of a mammal of a second species; (3) a transgenic mammal containing an artificial or a chimeric gene of the invention such as a transgenic mouse including a chimeric PrP gene wherein a portion of the mouse gene is replaced with a corresponding portion of a human PrP gene thereby rendering the mouse susceptible to infection with human prions; (4) a method of determining whether a sample is infected with prions which method involves inoculating a transgenic mammal of the invention with a sample to be 20 tested and observing the mammal for a period of time sufficient to determine if the mammal develops symptoms of a disease normally associated with prions; (5) a method of testing the efficacy of a drug in the treatment of disease developed as a result of infection with prions comprising administering a drug to be tested to a transgenic animal infected with prions and observing and/or testing the mammal to determine if the drug aids in promoting or slowing the progress of the disease or the symptoms; and (6) a method for determining the cause 30 of death of an animal comprising inoculating a transgenic animal of the invention with extracted brain tissue from the animal which has died and observing the transgenic animal in order to determine if the transgenic animal develops symptoms of prion infections.

Preferred host animals are mice and rats, with mice being most preferred in that there exists considerable knowledge on the production of transgenic animals. Other possible host animals include those belonging to a make selected from Mus, Rattus, Oryctolagus, and Mesocricetus. The host PrP gene can include codon from genetically diverse PrP genes from animals belonging to a genus selected from Bos, Ovis, Sus and Homo. Preferably, a mouse host PrP gene is changed to include a codon from a human, cow or sheep PrP gene, with human being most preferred.

The genetic material which makes up the PrP gene is 10 known for a number of different species of animals. Further, there is considerable homology between the PrP genes in different mammals. For example, see the amino acid sequence of mouse PrP compared to human, cow and sheep PrP in Figures 3, 4 and 5 wherein only the 15 differences as shown. Further, note that the segment of a PrP gene used to create the MHu2M gene of the present invention will result in encoding of protein which shows a difference between the human and a mouse protein of only nine residues. Although there is considerable genetic homology with respect to PrP genes, differences are significant in some instances. specifically, due to small differences in the protein encoded by the PrP gene of different mammals, a prion which will infect one mammal (e.g. a human) will not 25 normally infect a different mammal (e.g. a mouse). to this "species barrier", it is not generally possible to use animals such as mice as test animals in order to determine whether a particular sample contains prions which would normally infect a different species of anima! The present invention solves this such as a human. problem in a surprising manner.

At first, it might appear that useful transgeric test animals could be produced by substituting the FrF gene of the animal which would act as the test animal (e.g., a mouse) with the PrF gene of a different animal (e.g., a humar) and use the test animal to determine if it becomes infected with prions of the other animal i.e.

prions which normally inject only a human. However, when such transgenic animals are produced and used as test animals, they are not readily infected with the prions which normally infect the mammal from which their transgenic gene was taken. Thus, artificial or chimeric genes of the invention will not have all of the host codons replaced with different codons from the genetically diverse animal. A more specific description of how the species barrier was broken in accordance with the present invention is provided below.

Species Barrier Broken

The transmission of human CJD to apes and monkeys 1.5 - 3 years after intracerebral inoculation provided considerable interest in the causes of neurodegenerative diseases [Gibbs, Jr. et al., <u>Science 161:388-389 (1968)</u>]. Humans are not genetically diverse from apes and monkeys which accounts for the cross-species infectivity, although with a long incubation time. While the high cost of caring for nonhuman primates prevented extensive scudies of the human prion diseases, the transmissibility of these diseases stimulated studies of the animal prion analogues in rodents [Manuelidis et al., <u>Proc. Natl. Acad. Cci. USA 75:3422-3436 (1978); Manuelidis et al.</u>, <u>Proc. Natl. Proc. Natl. Acad. Sci. USA 73:223-227 (1976); Tateishi et al., <u>Ann. Neurol.</u> 5:581-584 (1979)].</u>

The present disclosure opens a new frontier in the investigation of the human prion diseases since transmission studies can now be performed relatively applied in genetically altered mammals such as Tg(MHu2M) mice that are relatively inexpensive to maintain. For the first time, endpoint titrations of prions in multiple human body tissues and fluids can be performed and standard curves constructed for more economical incubation time assays. The information derived from such studies of human prions will be useful in the management of CJD patients who are thought to pose some

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risk to relatives, physicians, nurses and clinical laboratory technicians [Berger et al., Neurology 43:205-206 (1993); Ridley et al., Lancet 341:641-642 (1993)].

In studies of human prion digeases with apes and 5 monkeys, the use of one or two, or rarely three, animals as recipients for a single inoculum has presented a significant problem in evaluating the transmissibility of a particular inoculum from an individual patient. Tg(MHu2M) mice described here obviate many of the 10 problems created by using nunhuman primates. The results demonstrate the "universality" of the MHu2M transgene for transmission studies with other types of transgenic animals and other prion inocula. For example, it may be most efficient to use mice expressing MHu2MPrP transgenes 15 coding for either a methionine or valine at codon 129, and by doing so, match the genotype of the Tg mouse (with respect to codon 129) with the genotype of the individual from which the inoculum is derived. Homozygosity at the codon 125 polymorphism has a profound influence on the 20 incidence of sporadic CJD [Palmer et al., Nature 352:340-342 (1991)]. The MHu2MPrP transgene encodes a Met at codon 129 and the iatrogenic CJD case was homozygous for Met [Collinge et al., Lancet 337:1441-1442 (1991)].

To break the species barrier we have created an artificial PrP gene which, when inserted into a first mammal (such as a mouse) renders that mammal susceptible to infection with prions which normally infect only a genetically diverse mammal (e.g. a human, cow or sheep). The artificial PrP gene may include the natural PrP gene sequence of the first animal with one or more (preferably less than 50) codon sequences being replaced with other codon sequences such as corresponding codons of a genetically diverse mammal (e.g. a human, cow or sheep).

In a specific example of the invention the species barrier is broken by inserting into a mammal (a mouse) the chimeric gene (MHu2M) which chimeric gene which is shown being assembled schematically in Figure 1. In

order to produce the chimeric gene, it is first necessary to obtain nucleotide sequences which encode human PrP. The human PrP genes are then subjected to conventional PCR procedures in order to produce large 5 numbers of copies of the gene or portions of the gene. The PCR product is then isolated, specific restriction sites are added and the copied product is subjected to specific endonucleases in order to remove a middle section of the human PrP gene. Specifically, restriction 10 sites are added such that when the PCR product is subjected to endonucleases such as Asp718 as well as BstEII, a section of the gene is cut out. these two endonucleases will remove a center portion of the human PrP gene (codons 94-188) which portion encodes 15 amino acid residues 94 through 188. Endonucleases are also used to remove a corresponding center portion of the mouse PrP gene. The removed center portion of the mouse gene is then discarded and the center portion obtained from the human PrP gene is fused into the mouse gene to 20 produce a chimeric human/mouse gene. Details of how the specific MHu2M gene was produced are described in Example 1 and shown in Figure 1.

As shown with Figure 2, there is a high degree of homology between the removed center portion of the human 25 PrP gene and the segment of the mouse PrP gene which is Specifically, the segments defer at nine replaced. Thus, when the genetic material is expressed, the resulting chimeric MHu2M protein will differ from MoPrP at 9 residues. These residues and their positions After the chimeric gene is 30 are shown in Figure 2. produced, it can be microinjected into a mouse egg using known technology as described within Scott et al., Cell 59:847-857 (1989) and Scott et al., Protein Sci. 1:986-997 (1992) and see also WO91/19810 published December 22, 1991 as well as other publications relating to the production of transgenic mice cited therein and known to those skilled in the art. The injected mouse egg is then

implanted into a mouse using known procedures. Multiple eggs can be implanted into a single mouse and known procedures can be used to determine whether the resulting offspring are transgenic mice which include the chimeric gene within their genome. Details of this procedure are described in Example 1.

It is not ; ossible to render a mammal susceptible to infection with prions from a genetically diverse mammal by substituting, completely, the PrP gene of the mammal 10 with the complete PrP gene of the genetically diverse mammal. We have successfully broken the "species barrier" by producing a chimeric PrP gene wherein a middle portion of the mouse PrP gene is replaced with a corresponding middle portion of a human PrP gene thereby 15 leaving the C- and N-terminus of the mouse PrP gene However, other segments of the mouse PrP gene can be replaced with other homologous segments of the human PrP gene and obtain a transgenic mouse which is subject to being readily infected with human prions. 20 Thus, the invention is not limited to the particular chimeric gene MHu2M or chimeric mice produced using this The invention includes all types of transgenic animals which include artificial genes wherein the artificial gene renders the transgenic animal susceptible 25 to infection with prions which normally infect only a genetically diverse animal.

Numerous specific examples of artificial genes of the invention can be deduced from reviewing Figures 3, 4 and 5. Specifically, one may start with the basic PrP gene of a mouse (as the first animal) which animal is to become the transgenic animal. Thereafter, one or more codons of the mouse gene may be replaced with one or more corresponding codons of a human, bovine or sheep PrP gene which codon is different from the corresponding codon of the mouse gene but at the same relative position in the gene. By showing that it is possible to break the "species barrier" by creating a particular chimeric gene

whereby transgenic mice can test for the presence of human prions we have opened the door for the creation of other transgenic animals which will include other artificial PrP genes which, for example, can allow for the testing for the presence of bovine or ovine prions in a sample.

Tq(MHu2M) mice with shorter incubation times

The incubation time of Tg(MHu2M) mice inoculated 10 with Hu prions is now about 200 days £50 days. We believe this can be reduced substantially. In Tg(SHaPrP) mice, the level of SHaPrP transgene expression was found to be inversely proportional to the length of the scrapic incubation time after inoculation with SHa prions [Prusiner et al., Cell 63:673-686 (1990)]. Thus, producing Tg(MHu2M) mice with higher levels of transgene expression should substantially reduce the incubation times.

Based on other studies which we have performed using 20 an analogous hamster/mouse chimeric PrP gene, MH2M, it is possible to assign a theoretical optimal incubation period for the MHu2M construct in a mouse lacking the endogenous mouse PrP gene. We obtained incubation periods of ~105 days with a heterologous Syrian hamster 25 prion inoculum, shortening to ~62 days with a homologous MH2M prion inoculum. The shortest incubation period so far observed in any of our transgenic mouse studies was ~45 days for a line expressing the mouse PrP gene. Assuming a similar minimum incubation period with MHu2M 30 prions in Tq(MHu2M PrP) mice lacking the endogenous mouse PrP gene, we can confidentially expect incubation periods of the order of $45 \times 105/62 = 76$ days with human prions. This is a conservative estimate, even shorter incubation periods may be possible in lines with very high copy numbers. Copy numbers can be increased up to 300-400 and the incubation timer can be as short as 50 days : 20 In addition, other chimeric human/mouse PrP days.

constructs may exhibit even shorter incubation times than MHu2M PrP.

In addition, removing MoPrP^c by crossing Tg(MHu2M) mice onto an ablated background (Prn-p^{0.70}) may also reduce the incubation time since Tg(SHaPrP^{*/0})81/Prn-p^{0.70} mice exhibit a 30% reduction in incubation times compared to Tg(SHaPrP*^{/0})81/Prn-p*^{/*} mice [Büeler et al., Cell 73:1339-1347 (1993). Prusiner, S.B. et. al., Proc. Natl. Acad. Sci. USA 90:10608-10612 Nov. 1993. Accordingly, we have also used fertilized eggs from mice in which the endogenous PrP gene has been ablated as recipients for microinjection of the MHu2M PrP construct.

By systematically altering the extent and position of the substitutions in other chimeric Hu/Mo PrP constructs, it is possible to further enhance the susceptibility of Tg mice to Hu prions as reflected by shortened incubation times. Shortening the incubation time is a worthwhile goal for the facilitation of many future studies in prion research and for the evaluation of pharmaceuticals.

In general, there is an inverse relationship between the number of copies of a chimeric or artificial PrP gene and the incubation time of disease after inoculation of the transgenic animal with prions. Specific 2Hu2M mice disclosed herein have only 3 or 4 copies of the 2Hu2M gene. The number of copies can be increased to 3 to 4 hundred thereby significantly reducing the incubation time from about 200 days to 50 days ± 20 days.

Differences in the conversion of MHu2MPrpc and HuPrpc into the scrapie isoform in mice

The fundamental event in prion propagation seems to be the conversion of PrP^c, which contains -43% α-helix and is devoid of β-sheet, into PrP^{sc} which has -43% β-sheet [Pan et al., Proc. Natl. Acad. Sci. USA 90:10962-10966 (1993)]. From the results of Tg(SHaPrP) mouse studies, this process is thought to involve the formation of a

complex between PrPsc and the homotypic substrate PrP [Prusiner et al., Cell 63:673-686 (1990)]. Attempts to mix PrPsc with PrPc have failed to produce nascent PrPsc [Raeber et al., J. Virol. 66:6155-6163 (1992)], raising the possibility that proteins such as chaperones might be involved in catalyzing the conformational changes that feature in the formation of PrPsc. One explanation for the difference in susceptibility of Tg(MHu2M) and Tg(HuPrP) mice to Hu prions in mice may be that mouse chaperones catalyzing the refolding of PrPc into PrPsc can recognize MHu2MPrP but not HuPrP.

Another possibility is that sequences at the N- or C-terminus of HuPrP inhibit the formation of PrPsc in murine cells. To test this possibility, HuPrP sequences are substituted for the Mo sequences at each terminus of MHu2MPrP. Comparison of the PrP sequences in many mammals around the GPI anchor addition site (codons 227-235) reveals an interesting difference of four amino acids between rodents and primates [Stahl et al., Biochemistry 31:5043-5053 (1992)]. In support of this hypothesis is that rodents also differ from ruminants including sheep and cattle at this site; sheep prions have failed to transmit neurodegeneration to Tg(ShePrP). In these experiments the entire mouse PrP gene was replaced with the entire sheep PrP gene.

Tg (MHu2M) contrast to mice, the transmission rate of Hu prion inocula from a wide variety of sources was less than 10% in Tg(HuPrP) mice, no different from the rate observed in non-Tg mice. 30 Likewise the conversion of HuPrpc into HuPrpsc in Tg(HuPrp) mice appears to be a relatively infrequent event similar to the rare conversion of $MoPrP^c$ to PrP^{sc} in response to The low rates of transmission in these human prions. mice do not seem to be a consequence of low titers of 35 human prion titers: two inocula which failed to cause disease in Tg(HuPrP) mice transmitted to 100% of inoculated Tg(MHu2M) animals.

"Strains" of human prions

Studies in rodents have shown that prion strains produce different patterns of PrPSc accumulation [Hecker et al., Genes & Development 6:1213-1228 (1992); DeArmond 5 et al., Proc. Natl. Acad. Sci. USA 90:6449-6453 (1993)); which can be dramatically changed by the sequence of Prp" [Carlson et al., Proc. Natl. Acad. Sci. USA in press (1994)]. The molecular basis of prion diversity has for many years been attributed to a scrapie specific nucleic 10 acid [Bruce et al., <u>J. Gen. Virol</u> 68:79-89 (1987)] but none has been found [Meyer et al., J. Gen. Virol. 72:37-49 (1991); Kellings et al., <u>J. Gen. Virol.</u> 73:1025-1029 Other hypotheses to explain prion strains include variations in PrP Asn-linked sugar chains [Hecker 15 et al., Genes & Development 6:1213-1228 (1992)] and multiple conformers of PrPsc [Prusiner, S.B., Science 252:1515-1522 (1991)]. The patterns of PrPSc in Tg(MHu2M) mice were remarkably similar for the three inocula from humans dying of CJD.

The patterns of PrPsc accumulation in the brains of inoculated Tq(MHu2M) mice were markedly different for RML However, RML prion Locula prions and Hu prions. containing MoPrPsc stimulated the formation of more MoPrPsc while Hu prion inocula containing HuPrpCJD triggered of MHu2MPrPsc. The distribution 25 production neuropathological changes characterized by neuronal vacuolation and astrocytic gliosis is similar to the patterns of PrPsc accumulation in the brains of Tg(MHu2M) mice inoculated with RML prions or Hu prions.

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New approaches to investigating human prion diseases

The remarkable sensitivity of Tg(MHu2M) mice to Hu important advance prions represents an neurodegenerative disease research. Based on the present 35 disclosure regarding chimeric Hu/Mo PrP transgenes we conceived of a similar approach to the construction of Tg mice susceptible to BSE and scrapie sheep priors. Such would be useful in detecting prion diseases in domestic animals. The importance of animal prion diseases is illustrated by BSE or "mad cow disease" in Great Britain, where >150,000 cattle have died. This prion disease BSE is thought to have originated with cattle consuming meat and bone meal produced from sheep offal containing scrapie prions [Wilesmith, J.W., Semin. Viro. 2, 239-245].

The BSE epidemic has led to considerable concern about the safety for humans of European beef and other cattle products. Epidemiologic studies over the past two decades have provided much data arguing that humans are unlikely to contract CJD from scrapie-infected sheep products [Harries-Jones et al., J. Neurol. Neurosurg. 15 <u>Psychiatry</u> 51:1113-1119 (1988); Cousens et al., Neurol. Neurosurg. Psychiatry 53:459-465 (1990); Brown et al., Neurology 37:895-904 (1987)]. There are seven amino acid substitutions which distinguish bovine from sheep PrP which must be considered in drawing conclusions 20 from sheep scrapie about the risk factors to humans from BSE. Whether any of these seven amino acid substitutions render bovine prions permissive in humans remains to be It may be that Tg(MHu2M) established. susceptible to bovine as well as sheep prions. Of 25 perhaps even greater importance, Tg(MHu2M) mice have immediate application in the testing of pharmaceuticals for human prion contamination. The (Hu 2M) described here provide a sensitive, reliable and economical bioassay for detecting the presence of human 30 prions.

Chimeric PrP Gene

Since the fundamental event underlying prion propagation seems to be a conformational change in PrP [Pan et al., <u>Proc. Natl. Acad. Sci. USA</u> 90:10962-10966 (1993)] and mouse PrP differs from human PrP at 31 positions out of 254 [Kretzschmar et al., <u>DNA</u> 5:315-324

(1986)], we constructed modified PrP transgenes. Chimeric SHa/Mo transgenes have produced prions with new properties, the most useful being the chimeric SHa/Mo transgene labeled MH2M which carries 5 amino acid substitutions found in SHaPrP lying between codons 94 and 188. [Scott et al., <u>Cell</u> 73:979-988 (1993)]. We made an analogous chimeric human/mouse PrP gene, which we call MHu2M, in which the same region of the mouse gene is replaced by the corresponding human sequence which differs from mouse PrP at 9 codons as is shown in Figure 2.

We have found that mice expressing the MHu2M chimeric transgene are susceptible to human prions after abbreviated incubation times. More specifically, the transgenic mice of the present invention which include the chimeric MHu2M gene will, after inoculation with human prions, develop disease symptoms attributed to the prions within about 200 days ± 50 days. Further, 80% or more the transgenic mice of the invention inoculated with human prions will develop symptoms of the disease, more preferably 98% or more of the mice will develop symptoms of the disease. According to experiments carried out, 100% of the transgenic MHu2M mice inoculated with human prions actually developed symptoms of the disease in about 200 days ± 50 days.

These findings indicate that murine cells cannot readily convert HuPrPc into HuPrPsc but they can process MHu2MPrPc into MHu2MPrPsc. Since Tg(MHu2M) mice develop neurodegeneration more rapidly than monkeys, they provide a preferred host for bioassays of infectivity in tissues of humans dying of prion diseases. The Tg(MHu2M) mice disclosed herein provide an excellent system for assessing the sterility of pharmaceuticals prepared from human sources. Other transgenic mice which include the prion protein gene of the animal in danger of infection can be used to test samples for prion diseases which can infect domestic animals such as sheep and cattle.

Chimeric MHu2M gone

Figure 1 is shown regarding how to breate the chimeric MHu2M gene. At first, we engineered a new Kpn1 site in the HuPrP ORF cassette (shown shaded), changing nucleotide 282 from a cytosine to a thymine residue by PCR-mediated mutagenesis. This mutagenic change conserves the amino acid sequence of HuPrP. A second oligonucleotide primer complimentary to DNA sequences around the BstEll-cut product was used to replace the corresponding MoPrP gene fragment (the MoPrP gene is unshaded) creating the hybrid gene MHu2M. Microinjection of a cosSHa.Tet construct bearing this expression cassette resulted in founder animal Tg(MHu2M)FVB-B5378.

An expanded representation of the region of MHu2MPr?

between codons 94 and 188 which is flanked by MoPi? sequences (Figure 2). MHu2MPrP differs from MoPrP by nine amino acids in the region between amino acids 96 and 167. These amino acid residues which are derived from HuPrP are shown on the lower section of the diagram; the amino acids at the same position of MoPrP are shown above. The discrepancy of amino acid positions is due to MoPrP having one less amino acid than HuPrP in the region immediately upstream from the replacement.

25 Artificial PrP Genes

The real power of the present invention lies in the understanding that a variety of different artificial PrP genes can be created which, when inserted into a first mammal, will render that mammal susceptible to infection with prions which normally only infect a second and genetically diverse mammal. There are nearly an infinite number of possible artificial PrP genes which would meet the basic criteria of the invention, i.e. rendering a mammal such as a mouse susceptible to infection with prions which normally infect only a genetically diverse mammal such as a human. The MHu2M gene of the invention is only one specific example of an artificial gene which

achieves the primary object of the invention. reviewing Figures 3, 4 and 5 numerous other artificial gene possibilities will be deduced by those skilled in Specifically, referring to Figure 3 one can 5 readily determine the amino acid sequence of mouse PrP and observe the positions wherein the mouse PrP sequence differs with a homan PrP sequence. Thus, to create an artificial gene one can substitute a codon (or sequence of codons) of a mouse PrP gene with a codon (or sequence 10 of codons) of a human PrP gene at the same position which will encode a different amino acid - any (but not all) of the codons where different sequences appear can be used for substitution. Peferring to Figure 4 it can be seen how it would be possible to produce artificial PrP genes 15 wherein the resulting gene could be inserted into a mouse in order to render the mouse susceptible to infection with bovine prions. A similar strategy with respect to producing a mouse which would be susceptible to infection with sheep prions can be deduced from reviewing Figure 5. 20 In addition to these possibilities those skilled in the art will recognize that, in certain instances, completely artificial nucleotide sequences can be corresponding substitutes for a portion of the natural sequence in order to obtain a useful artificial gene 25 which, when inserted into an animal, will render that animal susceptible to infection with prions which normally would infect only a genetically diverse mammal.

Evidence of Disease

Prp^{MC} has been found in the brains of affected Tg(MHu2M) mice after inoculation with Hu(CJD) or Mo(RML) prions. Brain homogenates of Tg(MHu2M) were either left undigested or digested with proteinase K (BMB) at a final concentration of 20 μg/ml for 1 hour at 37°C (even numbered lanes). Samples were resolved by SDSPAGE and then analyzed by Western blot.

The distribution of PrP⁵ and PrP^{5c} in clinically sick Tg(MHu2M) mice inoculated with Mo(RML) and Hu(CJD) prions were detected by the histoblot method. The histoblots included those of coronal sections through the region of the hippocampus and thalamus. Differences are observed between: (A) PrP^c in Mo(RML) infected mouse; (B) PrP^c in sporadic CJD RG-infected mouse; (C) PrP^{sc} in Mo(RML) infected mouse; (D) PrP^{sc} in sporadic CJD RG-infected mouse; (E) PrP^{sc} in sporadic CJD EC-infected mouse; and (F) PrP^{sc} in iatrogenic CJD (#364)-infected mouse.

EXAMPLES

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the chimeric genes, transgenic mice and assays of the present invention, and are not intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers used (e.g. amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is weight average molecular weight, temperature is in degrees Centigrade, and pressure is at or near atmospheric.

Example i

Construction of Chimeric gene (MHu2M)

The source of the HuPrP ORF for construction of an expression cassette has been described [Hsiao et al., Nature 338:342-345 (1989)]. The construction of the MHu2M gene is described in connection with the description of Figure 1. All PrP ORF cassettes were flanked by Sall and Xhol, which cleave immediately adjacent to the PrP initiation and termination codons of the PrP ORF respectively, allowing for convenient subcloning into the cos.SHaTet cosmid expression vector

[Scott et. al., <u>Cell</u> 73:979-988 (1993)]. The isolation and screening of recombinant cosmid clones was achieved by methods which have been proviously described [Scott et al., <u>Cell</u> 73:979-988 (1993)].

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Example 2

Producing Transgenic Mice/Tg(MHu2M)

The nucleotide sequences of the HuPrP and MHu2MPrP ORFs of Example 1 were verified. The cosmid Not1 10 fragments, recovered from large-scale DNA preparations, were used for microinjection into the pronuclei of fertilized C57BL/6 X SJL or FVB/N oocytes as previously described [Scott et al., Cell 59:847-857 (1989); Scott et al., Protein Sci. 1:986-997 (1992)]. Genomic DNA 15 isolated from tail tissue of weaning animals was screened for the presence of incorporated transgenes using a probe that hybridizes to the 3'-untranslated region of the SHaPrP gene contained in the cosSHa. Tet vector (Scott et al., Protein Sci. 1:986-997 (1992)]. The offspring 20 obtained were tested and it was confirmed that the chimeric MHu2M gene was integrated into the genome of these offspring. As shown in Example 5 below, these pice were found to be susceptible to infection with human prions 100% of the time.

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Example 3 Preparation of brain homogenates

A 10% [w/v] homogenate of a sample of thawed human brain tissue was prepared in phosphate buffered saline lacking calcium and magnesium ions. The tissue was initially dissociated using a sterile disposable homogenizer, and this suspension was subjected to repeated extrusion through an 18 gauge syringe needle followed by a 22 gauge needle. Samples for inoculation into test animals were diluted 10-fold. Homogenates of clinically sick Tg and non-Tg mouse brains were prepared

in the same way except for the omission of the initial dissociation step.

Example 4

5 Sources of prion inocula

Human inocula were derived from frozen brain tissues of patients in which the clinical diagnosis of CJD or GSS had been confirmed by histopathological examination of brain tissues and, in most cases, by prion protein analysis. In some cases, the PrP gene was amplified by PCR of DNA isolated from patient blood and the PrP sequence determined by DNA sequence analysis. No HuPrP mutations were detected in cases of sporadic or istrogenic CJD. The RML isolate was obtained from Swiss mice [Chandler, R.L., Lancet 1:1378-1379 (1961)] from a closed colony at the Rocky Mountain Laboratory or in Swiss CD-1 mice obtained from Charles River Laboratories.

Example 5

Determination of Scrapie Incubation Periods

Transgenic mice as per Example 2 were inoculated intracerebrally with 30 µl of brain extract using a 27 gauge needle inserted into the right parietal lobe. The preparation of inocula and criteria for diagnosis of scrapie in mice have been described [Carlson et al., Cell 46:503-511 (1986)]. Beginning 50 days after inoculation, the mice were examined for neurologic dysfunction every 3 days. When clinical signs of scrapie appeared, the mice were examined daily. When some animals whose death 30 was obviously imminent were identified, their brains were taken for histopathologic studies (as per the procedures of Example 3) and confirmation of the diagnosis of scrapie.

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Example 5

Immunoblot Analysis

For the determination of the relative levels of PrP expression in Tg mouse and human brains, protein concentrations were determined by bicinchoninic acid assay and immuno dots blots as previously described [Scott et al., Cell 73:979-988 (1993)]. Samples for Western blot analysis were prepared and western blots were performed as described previously [Towhin et al., Proc. Natl. Acad. Sci. USA 76:4350-4354 (1979)], except that an enhanced chemiluminescent (ECL) detection method (Amersham) was used. The blot was exposed to X-ray film for 5-60 seconds. α-PrP RO73 rabbit antiserum was used at a final dilution of 1:5000 and 3F4 monoclonal antibody was also employed [Serban et al., Neurology 40:110-117 (1990)].

Example 7

Tq(MHu2MPrP) Mice Are Susceptible to Human Prions

Using information from a systematic study of chimeric SHa/Mo PrP genes, a chimeric Hu/Mo PrP gene analogous to MH2M was constructed as per Example 1. This gene, which we call MHu2M, differs from MoPrP at 9 positions, all of which lie between codons 94 and 188 as is shown in Figure 2. A Tg mouse was constructed expressing the MHu2M PrP gene and the founder designated Tg(MHu2M)FVB-B5378 as per Example 2. By serial dilution of Tg(MHu2M) mouse and human brain homogenates, we estimate that the level of MHu2MPrPc in the brains of these Tg(MHu2M)FVB-B5378 mice are similar to those of HuPrPc found in human brain using an immuno dot blotting procedure.

Tg(MHu2M) mice as per Example 2 were inoculated with brain homogenates from three unrelated Caucasian patients who had died of CJD. Two of the three patients died of sporadic CJD: one (R.G.) was a 56-year-old American female; the other (E.C.) was a 61-year-old Canadian

female. In both cases, cerebrocortical biopsy showed severe spongiform degeneration. The third (#364) was a British youth who had contracted iatrogenic CJD after treatment for hypopituitarism with human growth hormone (HGH) derived from cadaveric pituitaries [Collinge et al., Lancet 337:1441-1442 (1991)]. Brain homogenates from all three CJD patients exhibited protease-resistant PrP by Western immunoblotting. This protease-resistant isoform of PrP is designated PrP^{SC}, or often PrP^{CJD} when it is found in humans.

All of the Tg (MHu2M) mice inoculated with homogenates from the CJD patients described above developed signs of central nervous system (CNS) dysfunction ~200 days after inoculation (see Table 1 The clinical signs were similar to those of After developing clinical signs, the murine scrapie. inoculated Tg(MHu2M) mice died rapidly, often within a Two of the eight uninoculated Tg(MHu2M) mice are now >500 days of age and remain well; the youngest of the 20 other six uninoculated animals is older than the age at which inoculated Tg(MHu2M) mice developed signs of CNS dysfunction.

Inoculation of Tg(MHu2M) mice with Mo(RML) prions passaged in mice produced disease in 178 ± 3 days, which is ~40 longer than Mo(RML) prions in non-Tg mice. Prolongation of incubation times in mice expressing non-murine transgenes is well established, and occurs presumably because the foreign PrP^c molecule inhibits the conversion of MoPrP^c into MoPrP^{sc} (Prusiner et al., Cell 63:673-686 (1990)). In contrast to Tg(MHu2M) mice, incubation times for RML prions in Tg(MH2M) mice were the same as those of the non-Tg mice (Scott et al., Cell 73:979-988 (1993)).

Table 1. Incubation of human (CJD) and mouse (RML) prion inocula Tg(MHu2M)FVB-B5378 mice	in

		Incut (mean	days ± SE)		
Source	Inoculum	No.	Range (days)	Illness	
Sporadic CID	RG	8/8	225-249		Death*
Sporadic CJD	EC	7/7		238±3.2	240±5.4(
latrogenic CJD	#364		202-229	218±4.6	N.D.
Мо		9/9	221-245	232 ± 2.9	235±3.9(
	RML	19/19	155-195	178±3.3	203±2.0(1

Number of animals developing clinical sickness (neurological dysfunction) divided by the total number of 15 animals inoculated. In the case of inoculum RG, three animals were found dead after 224, 238, and 243 days hefora a clinical diagnosis could be made. In the case of inoculum EC, two animals were found dead after 225 and 226 days before a clinical diagnosis could be made. 20 each case, these animals died at the same time that clinical diagnosis was made in other inoculated animals. The number of mice dying of scrapie is shown in parentheses. Mice sacrificed for pathological examination are excluded from these calculations. 25

Example 8

Comparative Example

Tg(HuPrP) Mice Are Resistant to Human Prions

Tg mice expressing HuPrP were produced using the HuPrP gene ORF, which had been cloned into the cosSHa. Tet expression vector [Scott et al., Protein Sci. 1:986-997 (1992)]. Microinjection of outbred C57B6/SJL and inbred FVB mouse embryos resulted in two founder transgenic animals designated Tg(HuPrP)B6SJL-110 and Tg(HuPrP)FVB-152. We estimated by serial dilution of brain homogenates and immuno dot blotting, that the level of PrPc in the brains of the progeny of these founders

express HuPrP at levels 4- to 8-fold higher than the level of HuPrP found in the human brain.

To determine whether expression of HuPrP Tq(HuPrP)B6SJL-110 and Tg (HuPrP) FVB-152 conferred 5 susceptibility to human prions, incubation periods were measured after inoculation of Tg(HuPrP) and non-Tg mice with brain extracts from 18 patients that had died of sporadic CJD, iatrogenic CJD, familial CJD or GSS. From experiments performed over the past 2.5 years, 10 concluded that the two lines of Tg(HuPrP) mice were no more responsive than non-Tg mice to human prions (see Table 2 below). The rate of transmission to Tq(HuPrP) mice was 8.3% (14 clinically sick mice out of 169 mice) which was similar to a transmission rate of 10.3% in 15 control non-Tg mice (6 clinically sick mice out of 58 In the infrequent event of a positive incubation times were extremely long transmission, ranging, from 590 days to 840 days in both Tg(HuPrP) and non-Tg mice. By this late time, many animals had died of 20 intercurrent illnesses which complicated diagnosis. difficulty of interpreting transmissions occurring after extremely long incubation periods is compounded by the heightened potential for artifactual results due to low levels of contaminating prions.

Statistical analysis shows that the frequency of Hu prion transmission to Tg (MHu2MPrP) mice compared to Tg (HuPrP) and non-Tg mice is highly significant using the Fisher's exact test, p<10⁻⁷ [Mehta et al., J. Am. Stat. Assn. 78: (392) 427-434 (1983)]. When Hu prion transmission to Tg (HuPrP) mice was compared to non-Tg mice, the frequencies were similar, p=0.79.

To confirm the clinical diagnosis of prion disease, 5 ill Tg(HuPrP) and 1 non-Tg mice were sacrificed and brain extracts were examined for the presence of PrPsc by Western blotting with the α-PrP antibodies, 3F4 mAb and RO73 antiserum [Kascsak et al., <u>J. Virol.</u> 61:3688-3693 (1987); Serban et al., <u>Neurology</u> 40:110-117 (1990)]. The

reacts specifically with HuPrP allowing discrimination from MoPrP. MoPrP' was detected in the brain of the non-Tq mouse inoculated with sporadic CJD inoculum #87011 which developed clinical signs after 756 5 days, while 3F4-reactive PrPsc was detected in the brains of two Tg(HuPrP) mice which developed clinical signs after 589 days post-inoculation with iatrogenic CJD inoculum #170. Notable is the finding that the proteaseresistant fragments of HuPrPsc from ill Tg(HuPrP) mice 10 migrate more rapidly on SDS-PAGE than do those of HuPrPGJP from human CJD brain and MoPrPsc from non-Tg mouse brain. Whether or not this reflects more extensive proteolysis of the N-terminus due to differences in conformation that result from altered folding of HuPrP in a mouse cell 15 remains to be established. The equivalent transmission rates of human prions in Tg(HuPrP) and non-Tg mice indicate that this is a rare event with the same frequency of occurrence as the stochastic conversion of MoPrPc to MoPrPsc induced by human prions.

The absence of either RO73- or 3F4-reactive PrPsc in the brains of 3 out of the 6 mice analyzed may reflect the difficulty of accurately diagnosing prion disease in elderly animals. Some of the mice inherited prion diseases of both humans and Tg mice exhibit little or 25 undetectable levels of protease-resistant PrP; yet, based on transmission studies, their brains contain prions and they show clear spongiform degeneration [Medori, R. et al, N. Engl. J. Med. 326, 444-449 (1992); Hsiao et al., (In preparation) (1994)].

In contrast to Tg(MHu2M) mice, Hu prions from 30 patient RG have not transmitted to either Tg(HuPrP) or non-Tg mice after >330 days (see Table 2 below). Attempts to transmit preparations enriched for Hu prion rods prepared from the brain of patient RG have likewise 35 been negative for >300 days. In addition, inoculum from the iatrogenic CJD case (#364) has produced illness in

neither Tg(HuPrP) nor non-Tg mice after >780 days (as shown in Table 2 below).

Table 2. Incubation times in Tg(HuPrP)FVB-152 and Tg(HuPrP)B6SJL-110 mice after inoculation with brain extracts from patients with human prion diseases

5	prion disease	S		
	Host	Inoculuin	(n/n _)"	Incubation times (days ± SE) ^b
_	Tg152	Sporadic CJD(#87011)	1/10	706
0,	Non-Tg	Sporadic CJD(#87011)	3/5	697.3±51
	Tg 152	Sporadic CJD(#88037)	3/10	680±28
	Tg 152	Sporadic CJD(RG)	0/10	
5	Non-Tg	Sporadic CJD(RG)	0/10	
	Tg 152	Sporadic (RG) Rods	0/8	
	Non-Tg	Sporadic (RG) Rods	0/8	
)	Tg 152	Codon 102 GSS(#87027)	4/10	724±16
	Non-Tg	Codon 102 GSS(#87027)	0/10	679
	Tg 152	Codon 102 GSS(#87031)	0/10	
5	Non-Tg	Codon 102 GSS(#87031)	1/5	742
	Tg 152	Codon 178 F-CJD	0/8	
	Non-Tg	Codon 178 F-CJD	0/8	
)	Tg 110	Sporadic CJD(#87036)	0/8	
	Non-Tg	Sporadic CJD(#87036)	1/5	838
	Tg 110	latrogenic CJD(#703)	0/10	
5	Non-Tg	latrogenic CJD(#703)	0/5	
	Tg 110	latrogenic CJD(#170)	2/10	589±0
	Non-Tg	latrogenic CJD(#170)	0/5	

	Tg 110	Iatrogenic CJD(#364)	0/10	
5	Non-Tg	Iatrogenic CJD(#364)	0/5	
	Tg 110	Codon 200 F-CJD	1/8	791
	Tg 110	Codon 217 GSS	1/8	874
l 0	Tg 110	Codon 102 GSS-A	0/10	
	Tg 110	Codon 102 GSS-B	1/8	694
	Tg 110	Codon 117 GSS	0/8	

- 15 * Number of animals developing clinical sickness divided by the total number of animals inoculated.
 - b Refers to time to diagnosis of illness.
 - c-f Patients from which inoculum were derived are described in the following publications: [Collinge et al., <u>Lancet</u>
- 20 337:1441-1442 (1991); Hsiao et al., <u>Nature</u> 338:342-345 (1989); Hsiao et al., <u>Neurology</u> 41:681-684 (1991)].

Example 9

Formation of MHu2MPrpsc or MoPrpsc In the Brains of Tg(MHu2M) Mice

Some clinically sick Tg(MHu2M) mice inoculated with each of the three CJD prion inocula or RML prions were sacrificed for his spathological verification of disease and for prion protein analysis. Western blots of brain homogenates from Tg(MHu2M) mice infected with Hu prions probed with RO73 and 3F4 \alpha-PrP antibodies revealed the presence of protease-resistant Prpsc which reacted with the 3F4 monoclonal antibody showing this protease-resistant product to be MHu2M Prpsc. The epitope recognized by this antibody consists of a pair of methionine residues at positions 109 and 112 in PrP [Rogers et al., J. Immunol. 147:3568-3574 (1991)] which

are contained in MHu2M but not in MoPrP as can be seen by the mouse/human comparison of figure 2. The polyclonal rabbit α-PrP antiserum RO73 diluted 1:5000 was poorly reactive with MHu2MPrPsc as well as HuPrPc and HuPrPsc from diseased human brains. Brain homogenates from Tg (MHu2M) mice infected with RML prions contained PrPsc which was detectable only with RO73 and not 3F4 α-PrP antibodies, indicating that Tg (MHu2M) mice are capable of producing MoPrPsc but not MHu2MPrPsc in response to RML prions previously passaged in mice. While these findings are similar to those reported for Tg(SHaPrP) mice (Scott et al., Cell 59:847-857 (1989)), they contrast with those found for Tg (MH2MPrP) mice where MH2MPrPsc was formed in response to RML prions [Scott et al., Cell 73:979-988 (1993)].

Example 10 Regional Distribution of Prpsc and Patterns of Neuropathology

The distribution of Mo and MHu2M PrPc and PrPsc is 20 shown in histoblots of coronal brain sections through the hippocampus and thalamus of Tg(MHu2M) mice inoculated with RML or CJD prions. The weak immunoreactivity of MHu2M PrP with R073 permitted a degree of analysis which 25 had not been previously possible in Tg mice expressing SHaPrP or MH2MPrP because these PrP species react with this antibody. The pattern of Prpsc deposition was highly dependent upon the species of origin of the infectious prions. When inoculated with RML prions, histoblots of 30 the brains of Tg(MHu2M) were similar to those of CD-1 mice infected with RML prions, revealing a diffuse pattern of MoPrPsc deposition in the hippocampus, thalamus, hypothalamus and all layers of the neocortex. The histoblot pattern of was strikingly different for Tq(MHu2M) mice inoculated with CJD prions. deposition of MHu2MPrPsc was confined primarily to the deep layers of the neocortex, the thalamus, particularly

the ventral posterior medial that in nucleus, the hypothalamus and the putamen. The hippocampal region and outer layers of the neocortex were spared. Interestingly, while the hippocampus was completely 5 devoid of MHulbler, this region showed the most intense MHu2MPrPc signal. The same pattern of MHu2MPrP30 deposition was consistently observed in histoblots of Tq(MHu2M) mice inoculated with all thre: CJD prion isolates prepared from human brain. It is noteworthy 1) that the pattern of MHu2MPrPsc deposition is similar to the pattern of P-PCUD deposition observed in histoblots of the brain from which inoculum RG was derived [Taraboulos et al., Proc. Natl. Acad. Sci. USA 89:7620-7624 (1992)]. The spongiform degeneration in the brains of Tg(MHu2M) 15 mice infected with Mo(RML) and Hu(CCD) prions reflected the patterns of PrPsc accumulation described above.

The instant invention is shown and described herein in what is considered to be a most practical and preferred embodiments. It is recognized, however, that departures may be made from which are within the scope of the invention and that modifications will occur to one who is skilled in the art upon reading this disclosure.

SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: The Regents of the University of California
 - (ii) TITLE OF INVENTION: METHOD OF DETECTING PRIORS IN A SAMPLE AND TRANSGENIC ANIMAL USED FOR SAME
 - (iii) MUNBER OF SEQUENCES: 4
 - ((v) CORRESPONDENCE ADDRESS:

 - (A) ADDRESSEE: Robbins, Bordiner & Carson (B) STREET: 201 N. Figueroa Street, 5th Floor
 - (C) CITY: Los Angeles
 - (C) STATE: CA
 - (E) COUNTRY: USA
 - (F) ZIP: 90012-2628
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk (8) COMPUTER: ISH PC competible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: Patentin Release #1.0, Version #1.25
 - (VI) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
 - (viii) ATTORNEY/AGENT INFORMATION:

 - (A) MANE: Berliner, Robert (B) REGISTRATION NUMBER: 20,121 (C) REFERENCE/DOCKET NUMBER: 5555-313
 - (1x) TELECOMPUNICATION INFORMATION:
 - (A) TELEPHONE: (213) 977-1001
 - (B) TELEFAX: (213) 977-1003
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 254 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: Linear
 - (ii) MOLECULE TYPE: peptide
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: HOUSE PRION PROTEIN, MOPEP
 - (x1) SEQUENCE DESCRIPTION: SEQ 10 NO:1:
 - Met Ala Asn Leu Gly Tyr Trp Leu Leu Ala Leu Phe Val Thr Met Trp 1 5 10 15
 - The Asp Vat Gly Leu Cys Lys Lys Arg Pro Lys Pro Gly Gly Trp Asn 20 25 30
 - The G y Gly Ser Arg Tyr Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg 35 40 45
 - Tyr Pro Pro Gin Gly Gly Thr Trp Gly Gin Pro His Gly Gly Gly Trp 50 60
 - Gly Gln Pro His Gly Gly Ser Trp Gly Gln Pro His Gly Gly Ser Trp 65 70 75 80
 - Gly Gln Pro His Sly Gly Gly Trp Gly Gln Gly Gly Gly Thr His Asn 85 90 95

Gln Trp Asn Lys Pro Ser Lys Pro Lys Thr Asn Leu Lys His vai Aia 100 105 110

Gly Ala Ala Ala Gly Ala Val Val Gly Gly Leu Gly Gly Tyr Met 115 120 125

Leu Gly Ser Ala Met Ser Arg Pro Met 11e His Phe Gly Asn Asp Trp 130 135 140

Glu Asp Arg fyr fyr Arg Glu Ash Met Tyr Arg Tyr Pro Ash Gln Val 145 150 155 160

Tyr Tyr Arg Pro Val Asp Gin Tyr Ser Asn Gin Asn Asn Phe Val His 165 170 175

Asp Cys Val Asn Ite Thr Ite Lys Gin His Thr Val Thr Thr Thr Thr 180 185 190

Lys Gly Glu Asn Phe Thr Glu Thr Asp Val Lys *** Met Glu Arg Val 195 200 205

Val Glu Rin Het Cys Val Thr Gin Tyr Gin Lys Glu Ser Gin Ala Tyr 210 215 220

Tyr Asp Gly Arg Arg Ser Ser Ser 1hr Vol Leu Phe Ser Ser Pro Pro 225 230 235 240

Val lie teu teu lie Ser Phe teu lie Phe teu lie Val Gly 245 250

(2) INFORMATION FOR SEG 10 NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 253 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: Linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: HUMAN PRION PROTEIN, HUPPP
- (xi) SEQUENCE DESCRIPTION: SEG 10 NO:2:

Het Ala Asn Leu Gly Cys Trp Het Leu Val Leu Phe Val Ala Thr Trp 1 5 10 15

Ser Asp Leu Gly Leu Cys Lys Lys Arg Pro Lys Pro Gly Gly Trp Ash $20 \\ 25 \\ 30$

The Gly Gly Ser Arg Tyr Pro Gly Gln Gly Ser Pro Gly Gly Ash Arg

Tyr Pro Pro Gin Gly Gly Gly Gly Trp Gly Gin Pro His Gly Gly Gly 50 60

Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly 65 70 75 80

Trp Gly Gin Pro His Gly Gly Gly Trp Gly Gin Gly Gly Gly the His 85 90 95

Ser Gin Trp Asn Lys Pro Ser Lys Pro Lys Thr Asn Met Lys His Met 100 105 110

Ala Gly Ala Ala Ala Ala Gly Ala Val Val Gly Gly Leu Gly Gly Tyr 115 120 125 Met Leu Gly Ser Ala Met Ser Arg Pro 11e 11e 31is Phe Gly Ser Asp 130 135 140

Tyr Glu Asp Arg Tyr Tyr Arg Glu Asn Met His Arg Tyr Pro Asn Gln 145 155 160

Val Tyr Tyr Arg Pro Met Asp Glu Tyr Ser Asn Gln Asn Asn Phe Val 165 170 175

His Asp Cys Val Asm Ile Thr Ile Lys Gln His Thr Val Thr Thr 180 185 190

The Lys Gly Glu Asn Phe The Glu The Asp Val. Lys Het Het Glu Arg $195 \hspace{1cm} 200 \hspace{1cm} 205 \hspace{1cm}$

Val Val Glu Gln Met Cys lie Thr Gln Tyr Glu Arg Glu Ser Gln Ala 210 215 220

Tyr Tyr Gln Arg Gly Ser Ser Met Val Leu Phe Ser Ser Pro Pro Val 225 230 240

lie Leu teu ile Ser Phe Leu lie Phe Leu ile Val Gly 245 250

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 263 amino acids
 - (8) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (34) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: BOVINE PRION PROTEIN, BOPEP
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Val Lys Ser His 11e Gly Ser Trp Ile Lou Val Leu Phe Val Ala 1 10 15

Het Trp Ser Asp Val Gly Leu Cys Lys Lys Arg Fro Lys Pro Gly Gly 20 25 30

Trp Asn Thr Giy Gly Ser Arg Tyr Pro Gly Gln Gly Ser Pro Gly Gly 35 40 45

Asn Arg Tyr Pro Pro Gln Gly Gly Gly Gly Trp Gly Gln Pro His Gly 50 55 60

Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly 65 70 75 80

Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly 85 90 95

Gly Gly Gly Trp Gly Gln Gly Gly Thr His Gly Gln Trp Asa Lys Pro 100 105 110

Ser Lys Pro Lys Thr Acn Met Lys His Val Ala Gly Ala Ala Ala Ala 115 120 125

Sty Ala Vat Vat Gty Gty Leu Gty Gty Tyr Met Leu Gty Ser Ala Met 130 135 140

Ser Arg Pro Leu Ile His Phe Gly Ser Asp Tyr Glu Asp Arg Tyr Tyr 145 150 155 160

Arg Giu Ash Met His Arg Tyr Pro Ash Glh Val Tyr Tyr Arg Pro Val 165 170 175

Asp Gin Tyr Ser Ash Gin Ash Ash Phe Val His Asp Cys Val Ash Ite 180 185 190

The Val Lys Glu His The Val The The The The Lys Gly Glu Asn Phe 195 200 205

The Glu The Asp Sie Lys Met Het Glu Arg Val Val Glu Gin Het Cys 210 215 220

Vel Thr Gin Tyr Gin Lys Giu Ser Gin Ale Tyr Tyr Asp Gin Gly Ale 225 230 235 240

Ser Val IIe Leu Phe Ser Ser Pro Pro Val IIe Leu Leu IIe Ser Phe 245 250 250

Leu lie Phe Lau Jie Val Gly 260

(2) INFORMATION FOR SEQ 10 NO:4:

- (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 255 amino acida
 - (8) TYPE: amino scid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: Linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL L'URCE-
 - (A) ORGANIAM: SHEEP PRION PROTEIN, ShPrP
- (ni) SEQUENCE DESCRIPTION: SEQ 10 NO:4:

Met Val Lys Ser His Ile Gly Ser Trp Ile Leu Val Leu Phe Val Ala 1 5 10 15

Met Trp Ser Asp Val Gly Leu Cys Lys Lys Arg Pro Lys Pro Gly Gly 20 25 30

Trp Asn The Gly Gly See Arg Tyr Pro Gly Gln Gly See Pro Gly Gly 35 40 45

Asn Arg Tyr Pro Pro Gln Gly Gly Gly Gly Trp Gly Gln Pro His Gly 50 60

Gly Gly Trp Gly Gin Pro His Gly Gly Gly Trp Glv Gin Pro His Gly 65 70 75 an

Gly Ser Trp Gly Gtn Pro His Gly Gly Cly G'y Trp Gly Gln Gly Gly 95

Ser His Ser Gin Trp Asn Lys Pro Ser Lys Pro Lys Thr Asn Met Lys 100 105 110

His Val Alm Gly Atm Atm Atm Atm 51y Atm Val Val Gly Gly Leu Gly 115 120 125

Gly Tyr Met Leu Gly Ser Ale Het Ser Arg Pro Leu Ile His Phe Gly 130 135 140

Asn Asp Tyr Glu Asp Arg Tyr Tyr Arg Glu Asn Met Tyr Arg Tyr Pro 145 150 155 160

Ash Gin Vel Tyr Tyr Arg Pro Ve. Asp Gin Tyr Ser Ash Gin Ash Ash 165 170 '75

Phe Vat His Asp Cys Vat Ash Ite Thr Vat Lys Gln His Thr Val Thr 180 - 185 - 190

The The Lys Gly Glu Asn Phe The Glu The Asp Ile Lys Ile Het 195 200 205

Gtu Arg Val Val Glu Gln Het Cys Ite Thr Gln Tyr Gln Arg Cth Sec 210 215 220

Gln Ala Tyr Tyr Gln Arg Glv Ala Ser Val Ile Leu Phe Ser Ser Pro 225 230 235 240

Pro Val Tie Leu Leu lie Ser Phe Leu ite Phe Leu Ile Val Gly 265 250 250

CLAIMS

- 1. An artificial PrP gene characterized such that when inserted into the genome of a first mammal 5 renders the mammal susceptible to infection with prions which normally infect only a genetically diverse second animal.
- 2. The artificial PrP gene of claim 1, 10 wherein the first mammal is a mouse or a rat and the second genetically diverse mammal is selected from the group consisting of a human, a cow and a sheep.
- 3. The artificial PrP gene of claim 1, wherein the gene is comprised of a natural codon sequence of a PrP gene of the first mammal with one or more, but not all, of its codons replaced with a different codon of a natural PrP gene of the second genetically diverse animal.

20

4. A chimeric gene, comprising:

C-terminus and N-terminus codon sequences of a PrP gene of a mammal of a first genus;

codon sequences of a PrP gene of a mammal of a second genus operatively connected between the N-terminus and C-terminus codon sequences from the mammal of the first genus, wherein the chimeric gene renders a mammal of the first genus susceptible to infection with a prion normally infectious only to a mammal of the second genus when the chimeric gene is operably inserted into the genome of a mammal of the first genus.

5. The chimeric gene of claim 4, wherein the first genus is selected from the group consisting of Mus. Rattus, Oryctolagus, and Mesocricetus.

- 6. The chimeric gene of claim 4, wherein the second genus is selected from the group consisting of Bos, Ovis, Sus, and Homo.
- 7. The chimeric gene of claim 4, wherein the mammal of the first genus is a mouse and the mammal of the second genus is a human.
- 8. A transgenic mammal having a genome comprised of an artificial PrP gene wherein the mammal is susceptible to infection with a prion which does not normally infect a species of the mammal lacking the chimeric PrP gene.
- 9. The transgenic mammal of claim 8, belonging to a genus selected from the group consisting of Mus, Rattus, Oryctolagus, and Mesocricetus.
- 10. The transgenic mammal of claim 8, wherein the artificial PrP gene comprises a portion of a PrP gene from a different genus of mammal.
- 11. The transgenic mammal of claim 10, wherein the different genus of mammal is selected from 25 the group consisting of Bos, Ovis, Sus, and Homo.
 - 12. The transgenic mammal of claim 11, wherein the different genus of mammal is of the species Homo sapiens.
 - 13. The transgenic mammal of claim 8, wherein the artificial gene is a mouse PrP gene having a portion thereof replaced with a human PrP gene.
- 35 14. The transgenic mammal of claim 13, wherein the artificial gene is MHu2M.

- 15. The transgenic mammal of claim 9, wherein the mammal has an 80% or greater chance of developing a disease as a result of being inoculated with a prion which does not normally infect a species of the mammal lacking the artificial PrP gene.
- 16. The transgenic mammal of claim 15, wherein the mammal has a 98% or greater chance of developing a disease as a result of inoculation with a prion which does not normally infect a species of the mammal lacking the artificial PrP gene.
- 17. The transgenic mammal of claim 8, wherein the mammal will exhibit a disease incubation time of about 300 days or less.
 - 18. The transgenic mammal of claim 17, wherein the incubation time is about 200 days \pm 50 days.
- 20 19. A method of testing a sample for the presence of prion, comprising:

inoculating a transgenic mammal with the sample, wherein the mammal has a genome comprised of an artificial PrP gene and is susceptible to infection with

25 a prion which does not normally infect a species of a mammal lacking the artificial PrP gene; and

observing the transgenic mammal in order to determine if the mammal develops symptoms of prion infection.

30

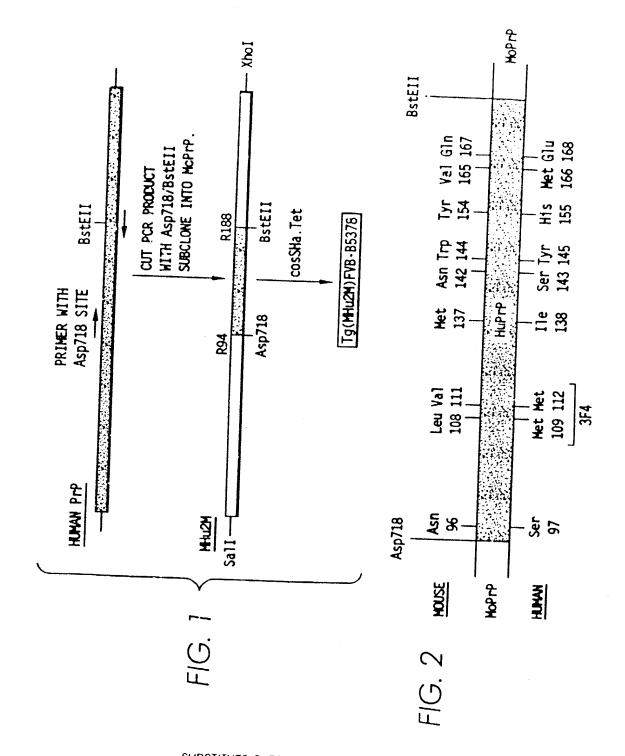
20. The method of claim 19, wherein the transgenic mammal is a mouse.

- 21. The method of claim 19, wherein the artificial gene is a chimer. The gene comprised of C-terminus and N-terminus codon sequences of the PrP gene of the mouse and, has fused operately thereto, codon sequences of a PrP gene of a human.
- 22. The method of claim 19, wherein the sample is a pharmaceutical formulation containing a therapeutically active component extracted from a human source.
- 23. The method of claim 22, wherein the pharmaceutical formulation is a formulation selected form the group consisting of injectables, orals, creams, suppositories, and intrapulmonary delivery formulations.
 - 24. The method of claim 19, wherein the sample is a cosmetic.
- 20 25. A method of determining the cause of death of an animal, comprising:

extracting brain tissue from an animal that has died;

inoculating a transgenic mammal with extracted brain tissue wherein the transgenic mammal has a genome comprised of an artificial PrP gene and is susceptible to infection with a prion from the animal that has died, which prion does not normally infect a species of a mammal lacking the artificial PrP gene; and observing the transgenic mammal in order

to determine if the transgenic mammal develops symptoms of prion infections.



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FIG. 3

to tu	Met	Ala	Asn	Leu	Gly	Tyr Cys	Trp	Leu Met	Leu	Ala Val	Leu	Phe	Val	Thr Ala		Trp	16
	Thr Ser	Asp	Val Leu	Gly	Leu	Cys	Lys	Lys	Arg	Pro	Lys	Pro	Gly	G1 y	Trp	Asn	32
lo lu	Thr	Gly	Gly	Ser	Arg	Туг	Pro	Gly	Gln	Gly	Ser	Fro	Gly	Gly	Asn	Arg	48
10 iu	Tyr	Pro	Pro	Gln	Gly	Gly		Thr Gly	Trp	Gly	Gln	Pro	His	Gly	Gly	Gly	63
lo lu	Trp	Gly	GIn	Pro	His	Gly	Gly	Ser Gly	Trp	G1y	Gln	Pro	His	Gly	G1 y	Ser Gly	79
lo lu	Trp	Gly	Gin	Pro	His	Gly	Gly	Gly	Trp	Gly	G1n	Gly	Gly	Gly	Thr	His	95
	Asn Ser	Gln	Trp	Asn	Lys	Pro	Ser	Lys	Pro	Lys	Thr	Asn	Leu Met	Lys		Val Met	111
lo iu	Ala	Gly	Ala	Ala	Aĭa	Ala	Gly	Ala	Val	Val	G1 y	G1y	Leu	Gly	GÌy	Tyr	127
to tu	Met	Leu	G1y	Ser	Ala	Met	Ser	Arg	Pro	Met Ile	Ile	His	Phe	Gly	Asn Ser	Asp	143
	Trp Tyr	Glu	Asp	Arg	Tyr	Tyr	Arg	G1u	Asn	Met	Tyr His	Arg	Туг	Pro	Asn	Gln	159
10 tu	Val	Tyr	Tyr	Arg	Pro	Val Met	Asp	G1n G1u	Tyr	Ser	Asn	G1n	Asn	Asn	Phe	Val	175
to tu	His	Asp	Cys	Val	Asn	Ile	Thr	Пe	Lys	Gln	His	Thr	Val	Thr	Thr	Thr	191
to tu	Thr	Lys	Gly	Glu	Asn	Phe	Thr	Glu	Thr	Asp	Val	Lys	Met	Met	Glu	Arg	207
Mo Hu	Va1	Val	G1u	Gln	Met	Cys	Val Ile	Thr	Gln	Tyr	Gln Glu		Glu	Ser	G1n	Ala	223
Mo Hu	Tyr	Tyr	_ '	Gly	Arg	Arg	Ser Gly	Ser	Ser	Thr Met	Val	Leu	Phe	Ser	Ser	Pro	239
Mo Hu	Pro	Val	Пе	Leu	Leu Su	Ile BSTIT						Leu	He	۷a۱	Gly		254

FIG. 4

Mo Bo	Met	Ala Val	Asn Lys	Leu Ser	His	 Ile	Gly	Tyr Ser	Trp	Leu Ile	Leu	Ala Val	Leu	Phe	Val	Thr Ala	14
Mo Bo	Met	Trp	Thr Ser	Asp	Val	Gly	Leu	Cys	Lys	Lys	Arg	Pro	Lys	Pro	Gly	G1 y	30
Mo Bo	•••	Trp	Asn	Thr	Gly	Gly	Ser	Arg	Tyr	Pro	G1 y	Gln	Gly	Ser	Pro	G1 y	4 5
Mo Bo	Gly	Asn	Arg	Tyr	Pro	Pro	Gln	Gly		 Gly		Trp	Gly	G1n	Pro	His	60
Mo Bo	Gly	Gly	Gly	Trp	G1y	Gln	Pro	His	Gly	Gly	Ser Gly	Trp	Gly	Gln	Pro	Hts	76
Mo Bo	Gly	G1y	Ser Gly	Trp	Gly	GIn	Pro	His	Gly	Gly	G1 y	Trp	Gly	Gln	 Pro		90
			 G!y					Gly	G1y	Thr	His	Asn Gly	Gln	Trp	Asn	Lys	100
Mo Bo	Pro	Ser	Lys	Pro	Lys	Thr	Asn	Leu Met	Lys	H1s	Val	Ala	Gly	Ala	Ala	Ala	116
Mo Bo	Ala	G1y	Ala	Val	Val	G1y	Gly	Leu	Gly	Gly	Tyr	Met	Leu	Gly	Ser	Ala	132
Mo Bo	Met	Ser	Arg	Pro	Met Leu	Ile	His	Phe	Gly	Asn Ser	Asp	Trp Tyr	Glu	Asp	Arg	Туг	148
Mo Bo	Tyr	Arg	Glu	Asn	Met	Tyr H1s	Arg	Tyr	Pro	Asn	Gln	Val	Tyr	Tyr	Arg	Pro	164
Mo Bo	Val	Asp	Gln	Tyr	Ser	Asn	Gln	Asn	Asn	Phe	Val	His	Asp	Cys	Val	Asn	180
Mo Bo	He	Thr	Ile Val	Lys	G1n G1u	His	Thr	Val	Thr	Thr	Thr	Thr	Lys	Gly	Glu	Asn	200
Mo Bo	Phe	Thr	Glu	Thr	Asp	Val Ile	Lys	Met	Met	Glu	Arg	Val	Val	Glu	Gln	Met	212
Mo Bo		Val	Thr	Gln	Tyr	G1n	Lys	Glu	Ser	Gln	Ala	Tyr	Tyr	Asp	Gly Gln		228
			Ser Ala	Ser		Val Ile	Leu	Phe	Ser	Ser	Pro	Pro	Val	Πe	Leu	Leu	244
Mo Bo		Ser	Phe	Leu		Phe UBST!											254

FIG. 5

Mo Met Ala Asn Leu --- Gly Tyr Trp Leu Leu Ala Leu Phe Val Thr 14
Sh Val Lys Ser His Ile Ser Ile Val Ala

Mo Met Trp Thr Asp Val Gly Leu Cys Lys Lys Arg Pro Lys Pro Gly Gly 30 Sh Ser

Mo --- Trp Asn Thr Gly Gly Ser Arg Tyr Pro Gly Gln Gly Ser Pro Gly 45 Sh

Mo Gly Asn Arg Tyr Pro Pro Gln Gly Gly · · · Thr Trp Gly Gln Pro His 60 Sh Gly Gly

Mo Gly Gly Gly Trp Gly Gln Pro His Gly Gly Ser Trp Gly Gln Pro His 76 Sh Gly

Mo Gly Gly Ser Trp Gly Gln Pro His Gly Gly Gly ··· Trp Gly Gln Gly 91 Sh

Mo Gly Gly Thr His Asn Gln Trp Asn Lys Pro Ser Lys Pro Lys Thr Asn 107 Sh Ser --- His Ser

Mo Leu Lys His Val Ala Gly Ala Ala Ala Ala Gly Ala Val Val Gly Gly 123 Sh Met

Mo Leu Gly Gly Tyr Met Leu Gly Ser Ala Met Ser Arg Pro Met Ile His 139 Sh Leu

Ho Phe Gly Asn Asp Trp Glu Asp Arg Tyr Tyr Arg Glu Asn Met Tyr Arg 155 Sh Tyr

Mo Tyr Pro Asn Gln Val Tyr Tyr Arg Pro Val Asp Gln Tyr Ser Asn Gln 171 Sh

Mo Asn Asn Phe Val His Asp Cys Val Asn Ile Thr Ile Lys Gln His Thr 187 Sh Val

Mo Val Thr Thr Thr Lys Gly Glu Asn Phe Thr Glu Thr Asp Val Lys 203 Sh Ile

Mo Met Met Glu Arg Val Val Glu Gln Met Cys Val Thr Gln Tyr Gln Lys 219 Sh Ile Arg

Mo Glu Ser Gln Ala Tyr Tyr Asp Gly Arg Arg Ser Ser Ser Thr Val Leu 235 Sh Gln --- Gly Ala Val Ile

Mo Phe Ser Ser Pro Pro Val Ile Leu Leu Ile Ser Phe Leu Ile Phe Leu 251 Sh

Mo Ile Val Gly

254

Sh

INTERNATIONAL SEARCH REPORT

Into Lational application to a
PCT/US95-04426

A. CLA	SSIFICATION OF SUBJECT MATTER										
IPC(6) C07H 15/12, C12N 15/00; A61K 49/00											
US CL 536/23.4; 800/2; 424/9 According to Internetional Patent Classification (IPC) or to both national classification and IPC											
		national classification and IPC									
Minimum documentation searched (classification system followed by classification symbols)											
	536/23.4; 800/2, 424/9	by classification symbols;									
Documentat	ion searched other than minimum documentation to the	extent that such documents are included	in the fields searched								
Electronic d	lets base consulted during the international search (na	me of data have and, where practicable	acarch terms used								
	EMICAL ABSTRACTS	, , , , , , , , , , , , , , , , , , ,									
C. DOC	L NTS CONSIDERED TO BE RELEVANT										
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